# In-Fusion<sup>™</sup> Dry-Down PCR Cloning Kit Protocol-at-a-Glance

## (PT3754-2)

Please read the *User Manual* (PT3754-1) before using this Protocol-at-a-Glance. This abbreviated protocol is provided for your convenience, but is not intended for first-time users.

### A. Cloning Procedure

- 1. Mix your PCR fragment and vector together at a 2:1 molar ratio in 10  $\mu$ l of deionized H<sub>2</sub>O.
- 2. Set up In-Fusion cloning reactions:
  - a. Cut one tube off the strip, and peel back the aluminum seal.
  - b. Add the 10 µl of vector + insert DNA (from Step 1). Mix well by pipetting up and down.
- 3. Incubate reactions at 42°C or at room temperature for 30 min, then transfer tubes to ice.
- 4. Proceed with Transformation (Section B). If you cannot transform cells immediately, store cloning reactions at –20°C until you are ready.

### **B.** Transformation

- 1. Dilute the In-Fusion reaction mixture with 40  $\mu$ I TE buffer, and mix well.
- 2. Transform competent cells with 2.5  $\mu$ l of diluted reaction mixture as follows:
  - a. Using Fusion-Blue<sup>™</sup> Competent Cells:
    - Thaw one vial of frozen Fusion-Blue Competent Cells on ice. Tap tube gently to ensure that the cells are suspended.
    - Add 2.5 µl of the diluted reaction mixture to the cells. Mix gently to ensure even distribution of the DNA solution. Leave the tube on ice for 30 min.
    - Do not add more than 5 µl of diluted reaction to 50 µl of competent cells.
    - Heat shock the cells in a water bath at 42°C for 45 sec, and then place them directly on ice for 1 min.
    - Competent cells should yield >1 x 10<sup>8</sup> cfu/µg. If not, replace with a fresh batch of cells.
  - b. If using other competent cells with In-Fusion™ Kits, follow the transformation protocol provided by the manufacturer (do not add more than 5 µl of diluted reaction to 50 µl of competent cells) and proceed to Step 3.
- 3. After heat shocking, add 450  $\mu l$  of SOC medium to the cells. Incubate at 37°C for 60 min while shaking at 250 rpm.
- 4. Take 1/20–1/10 of the cells (25–50 μl) from each transformation. Bring the volume up to 100 μl with SOC medium, and plate by spreading on separate LB/X-Gal/Amp plates. Spread the remaining cells from each transformation on separate, LB/X-Gal/Amp plates. Incubate all plates at 37°C overnight.
- 5. The next day, pick white colonies from each experimental plate and isolate plasmid DNA using a standard method of your choice.

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